Foliar diseases on tea and maté in Argentina caused by *Pseudocercospora* species

### Braun U¹, Rybak M², Rybak R² and Cabrera MG³

¹Martin-Luther-Universität, Institut für Biologie, Bereich Geobotanik und Botanischer Garten, Herbarium, Neuwerk 21 D-06099 Halle/S, Germany
²Instituto Nacional de Tecnología Agropecuaria (INTA), Estación Experimental Agropecuria, Cerro Azul, Misiones, Argentina
³Universidad Nacional del Nordeste, Facultad de Ciencias Agrarias, Sargento Cabral 2131, Corrientes, Argentina

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The leaf-spotting hyphomycete *Pseudocercospora theae* on tea (*Camellia sinensis*) is described from the Province Misiones in Argentina. This is the first record of this disease from Argentina. The taxonomy of *Pseudocercospora* species on *Camellia* spp. is briefly discussed, and the species concerned are keyed out. *Pseudocercospora mate* is an endemic cercosporoid hyphomycete on maté (yerba mate, *Ilex paraguariensis*) described from Argentina. This little known species is re-described on the base of new rich specimens recently collected in Misiones. The taxonomy of this species is discussed, above all the differentiation between *P. mate* and confused Asian collections that belong to *P. naitoi*. A key to *Pseudocercospora* species on *Ilex* spp. is provided.

**Key words** – Cercosporoid hyphomycetes – Misiones – *Pseudocercospora mate* – *Pseudocercospora theae*

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*Corresponding author: Uwe Braun – e-mail – uwe.braun@botanik.uni-halle.de*

### Introduction

Tea (*Camellia sinensis* (L.) Kuntze) and maté (yerba mate, Paraguay tea; *Ilex paraguariensis* A. St.-Hil.) are economically important crops in the Province Misiones of Argentina. Data on yield losses of these cultivated plants in Argentina caused by foliar diseases are old and scantly, and date back to 1983 in the case of maté (Rivera Flores 1983). Therefore, a research project on foliar diseases of these crops was carried out between 2009 and 2011. Several causal agents of foliar damages were identified, including cercosporoid hyphomycetes which were among the most important fungal pathogens due to the frequency of occurrence and initiation of premature defoliation. The leaf-spotting hyphomycete *Pseudocercospora theae*, found in Argentina for the first time, is described, illustrated and compared with other *Pseudocercospora* spp. on tea. The endemic *P. mate* is the cause agent of a foliar disease on maté. Based on new collections, this little known species is re-described, illustrated, compared with allied species, and a key to the species concerned is provided.

### Methods

The fungal material was examined by standard light microscopy using oil immersion (bright field and phase contrast), but without any staining. Thirty measurements (× 1000 magnification) of conidia and other structures were made, with the extremes given in parentheses.
Results and discussion

Figs 1–3

Leaf spots subcircular to somewhat angular or irregular, 1–10 mm diam., at first dark dingy greenish or greyish green, later pale to medium brown, greyish brown, finally grey to greyish white on the upper leaf surface and brown to greyish brown below, margin indefinite or with a narrow darker border, pale to mostly dark brown, blackish brown or purple, occasionally with a diffuse yellowish halo. Caespituli amphigenous, conspicuous on the upper side, scattered, punctiform, dark brown to blackish, less conspicuous below.

Mycelium internal; hyphae branched, septate, 1–4 µm wide, subhyaline to olivaceous, smooth. Stromata almost lacking to well-developed, epiphyllous stromata immersed, large, 40–100(–150) µm diam., circular to irregular in outline, hypophyllous stromata almost lacking or smaller, 10–50 µm diam., substomatal or occasionally intraepidermal, pale to dark olivaceous-brown, stoma cells 1.5–6 µm diam. Conidiophores in small, loose to large, dense fascicles, arising from stromata, on the upper side in large sporodochial conidiomata, erumpent, on the lower side in small to moderately large fascicles, emerging through stomata or occasionally erumpent through the cuticle, erect, straight to geniculate-sinuous, usually unbranched, occasionally with short, peg-like terminal branchlets, 5–40 × 1.5–4 µm, 0–1(–2)-septate, subhyaline to pale olivaceous or olivaceous-brown, thin-walled, smooth. Conidiogenous cells integrated, terminal, or conidiophores reduced to conidiogenous cells, 5–20 µm long; conidiogenous loci (scars) inconspicuous, sometimes denticle-like, neither thickened nor darkened. Conidia solitary, narrowly obclavate-subcylindrical, long conidia sometimes subacicular-filiform, short conidia occasionally cylindrical, (12–)20–100 × 1.5–4 µm, indistinctly 1–8-septate, hyaline, subhyaline or with a pale greenish or very pale olivaceous tinge, thin-walled, smooth, attenuated towards a subacute apex or tip subobtuse, base short to long obconically truncate, hilum neither thickened nor darkened, 1–1.5(–2) µm wide.

Fig. 1 – _Pseudocercospora mate_ on _Ilex paraguariensis_, based on HAL 2504 F. A Sporodochial fascicle of conidiophores. B Small fascicle of conidiophores emerging through a stoma. C Conidiophores. D Conidia. Bar: 10 µm. U. Braun del.
Material examined – **Argentina**, Misiones, Santa Anna, on leaves of *Ilex paraguariensis*, 20 June 1909, B. Zamboni (LPS, lectotype, designated by Braun 2000); Misiones, San Ignacio, on *I. paraguariensis*, August 1908, J. Torre (LPS, syntype, exhausted); Misiones, Aristóbulo del Valle, 17 Mar. 2011, M. Rybak (CTES 0566507, HAL 2504 F).

**Fig. 2** – *Pseudocercospora mate* on *Ilex paraguariensis*, based on fresh material. Epiphyllous leaf spots. Bar: 1 cm.

*Cercosporina mate*, described by Spegazzini (1910), was re-described by Chupp (1954) as *Cercospora mate*. Sutton & Pons (1980) examined syntype material of this species collected by J. Torre at San Ignacio, but without any conclusion about its generic affinity and taxonomy as this collection is devoid of any fructification, which was also confirmed by Braun (2000) who examined type collections of this species as well. However, the latter author found fungal colonies of *C. mate* in a second syntype collection that he designated as lectotype. Based on inconspicuous conidiogenous loci in the lectotype material, he concluded that this species has to be reallocated to *Pseudocercospora*, but meanwhile Guo & Zhao (1989) had introduced the new combination *Pseudocercospora mate* just based on Chinese collections, i.e. without comparing the Chinese samples with type material.

**Fig. 3** – *Pseudocercospora mate* on *Ilex paraguariensis*. Lesions on leaves in a plantation after premature fall. Bar: 1 cm.

Braun (2003) compared Japanese collections of *Pseudocercospora naitoi* (Nakashina et al. 1999) on *Ilex chinensis* Sims (= *I. oldhamii* Miq.) [CUP 40367] and *I. serrata* Thunb. [CUP 40747] with *P. mate* and found obvious morphological differences. He concluded that Chinese collections on *Ilex* spp., misidentified as *P. mate* by Guo & Zhao (1989), Guo & Hsieh (1995) and Guo et al. (1998), belong to *P. naitoi* as well. The latter species is distinguished from *P. mate*, which is confined to *Ilex paraguariensis* in South America, by its olivaceous conidia, much smaller stromata and the formation of superficial hyphae with solitary conidiophores which are present in Chinese as well as Japanese collections, although not described in Nakashina et al. (1999). The Asian *Pseudo-
*cercospora* species on *Ilex* spp. is characterized as follows

**Pseudocercospora naitoi** (Togashi) C. Nakash. & Tak. Kobay., Mycoscience 40: 272, 1999

Fig. 4


Fig. 4 – *Pseudocercospora naitoi* on *Ilex chinensis* (= *I. oldhamii*), based on CUP 40367. **A** Superficial hypha. **B** Superficial hyphae with solitary conidiophores. **C** Fascicles of conidiophores. **D** Conidiophores. **E** Conidia. Bar: 10 µm. U. Braun del.

Leaf spots amphigenous, subcircular to angular-irregular, sometimes confluent, 1–15 mm diam., reddish brown to blackish on the upper surface, greyish brown or grey below, margin indefinite or with a pale yellowish brown halo, sometimes spots greyish white with dark border. *Caespituli* amphigenous, punctiform, scattered, dark. *Mycelium* internal and partly also external, superficial; *hyphae* branched, 1.5–3 µm wide, septeate, subhyaline to pale olivaceous or olivaceous-brown, thin-walled, smooth. *Stromata* immersed or substomatal, 10–50 µm diam., olivaceous-brown, cells 1.5–4 µm diam. *Conidiophores* in small to moderately large fascicles, loose to rather dense, arising from stromata, emerging through stomata or erumpent, or solitary, arising from superficial hyphae, erect, straight, subcylindrical-filiform to moderately geniculate or sinuous, usually unbranched, length variable, sometimes uniformly short, sometimes short and long conidiophores mixed, 5–50(–65) × (1.5–)2–4(–5) µm, 0–3-septate, pale olivaceous or olivaceous-brown, darker in mass, thin-walled, smooth; *conidiogenous cells* integrated, terminal or conidiophores reduced to conidiogenous cells, 5–30 µm long, *conidiogenous loci* inconspicuous. *Conidia* solitary, obclavate to obclavate-cylindrical, (20–)30–85(–100) × (1.5–)2–3.5(–5) µm, 3–11-septate, pale olivaceous, thin-walled, smooth, apex subacute, base truncate to short obconically truncate, 1–2 µm wide, hilum unthickened, not darkened.

Naito (1940) cited Japanese collections as “Cercospora mate” (he did not intend to introduce a new combination based on *Cercosporina mate*, but in fact did so by accident).

There are some additional species of *Pseudocercospora* on *Ilex*, viz. the North American *P. ilicis* (Ellis) U. Braun (Chupp 1954: 53, Braun 2003: 85 and 95, Fig. 9), *P. ilicis-micrococcae* Goh & W.H. Hsieh (Hsieh & Goh 1990: 26 and 28, Fig. 14) endemic in Taiwan on *Ilex maximowicziana* Loes. and *I. micrococca* Maxim., the North American *P. pulvinula* (Cooke & Ellis) U. Braun (Chupp 1954: 54, Braun 2003: 86 and 95, Fig. 10), and the South American *P. yerbae* (Speg.) U. Braun (Chupp 1954: 55, Braun 2000: 74 and 75, Fig. 19). The following key contains all
Pseudocercospora species known to be leaf-spotting pathogens on Ilex spp.:

**Key to the species of Pseudocercospora on Ilex spp.**

1. Caespituli very conspicuous, punctiform, dark brown to blackish, sporodochial, stromata well-developed, very large, 30–200 µm diam., either very large on the upper leaf side and smaller below or large and well-developed on the lower side; on Ilex spp. in North and South America .................. 2

1* Caespituli less conspicuous, stromata either lacking or smaller, 10–50 µm diam.; on Ilex spp. in Asia, Europe or North America .... 4

2. On the upper leaf side with large pustulate sporodochial conidiomata, stromata very large, conidiophores very numerous, densely aggregated, on the lower side stromata lacking or small, conidiophores in smaller, looser fascicles or solitary, arising from superficial hyphae; on Ilex spp. in North America .................. *P. pulvinula*

2* Mycelium internal, superficial hyphae with solitary conidiophores lacking; on Ilex spp. in South America ........................................ 3

3. Epiphyllous caespituli very conspicuous, stromata immersed, large, 40–100(–150) µm diam., hypophyllous caespituli less conspicuous, stromata lacking or smaller, 10–50 µm diam., substomatal to intraepidermal; conidia (12–)20–100 × 1.5–4 µm, hyaline or subhyaline; on Ilex paraguariensis, South America (Argentina) .................... *P. mate*

3* Caespituli only hypophyllous, conspicuous, stromata always large, 75–200 µm diam.; conidia wider, 15–60 × 4–6 µm, pale olivaceous; on Ilex spp., South America (Argentina) ....................... *P. yerbae*

4. Conidiophores long and often branched, 45–155 µm; on Ilex spp., Asia (Taiwan) ........................................... *P. ilicis-micrococcae*

4* Conidiophores much shorter and/or unbranched ........................................ 5

5. Conidiophores short, 5–50(–65) × (1.5–)2–4(–5) µm, thin-walled; conidia (1.5–)2–3.5(–5) µm wide, hila 0.75–1.5 µm wide; superficial hyphae with solitary conidiophores present; on Ilex spp., Asia (China, Japan) .............................................. *P. naitoi*

5* Conidiophores longer and wider, (20–)30–90(–125) × 4–6 µm, wall somewhat thickened; conidia somewhat wider, 2–5 µm, hila (1.5–)2(–2.5) µm wide; superficial hyphae and solitary conidiophores lacking; on Ilex spp., North America ............ *P. ilicis*

**Pseudocercospora theae** (Cavara) Deighton, Trans. Brit. Mycol. Soc. 88: 390, 1987 Fig. 5

≡ *Septoria theae* Cavara, Rev. Mycol. 11(44): 190, 1889.


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**Fig. 5** – *Pseudocercospora theae* on *Camellia sinensis*, based on HAL 2505 F. A Sporodochial fascicles of conidiophores. B Small fascicle of conidiophores emerging through a stoma. C Conidiophores. D Conidia. Bar: 10 µm. U. Braun del.
Fig. 6 – *Pseudocercospora theae* on *Camellia sinensis*, based on fresh material. Epiphyllous leaf spots. Bar: 1 cm.

**Leaf spots** amphigenous, circular to somewhat angular-irregular, 0.5–3 mm diam., ochraceous, pale to medium dark brown, later centre becoming greyish brown, grey to greyish white, with narrow raised rim followed by a dark marginal line or halo. **Caespituli** amphigenous, punctiform, scattered, greyish brown to blackish. **Mycelium** internal, **hyphae** 1–2.5 µm wide, hyaline to pale olivaceous. **Stromata** lacking to well-developed, 10–60 µm diam., substomatal or intraepidermal (on the upper leaf side immersed, larger and well-developed, substomatal; lacking or smaller below). **Conidiophores** in small, loose to rich and dense fascicles (large fascicles with large stromata sporodochial), arising from internal hyphae or stromata, erumpent or emerging through stomata, erect, subcylindrical to strongly geniculate-sinuous, unbranched, rarely with short terminal branchlets, 5–40 × 1.5–4 µm, 0–2-septate, pale olivaceous, thin-walled, smooth; **conidiogenous cells** integrated, terminal or conidiophores reduced to conidiogenous cells, 5–25 µm long; **conidiogenous loci** inconspicuous, neither thickened nor darkened.

**Conidia** solitary, acicular, i.e. with truncate base and gradually attenuated towards a subacute tip, to narrowly obclavate, i.e. base obconically truncate and apex obtuse to subacute, short conidia occasionally sub-cylindrical, 30–90 × 1.5–4 µm, (1–)3–9-septate, hyaline, subhyaline to very pale olivaceous, thin-walled, smooth, 1.5–2(–2.5) µm wide at the truncate base, hila unthickened, not darkened.

Material examined – **Argentina**, Misiones, Aristóbulo del Valle, on *Camellia sinensis*, 17 Mar. 2011, M. Rybak (CTES 0566508, HAL 2505 F)

This is the first record of *Pseudocercospora theae* from Argentina. This species is also known from Africa (Malawi, Mauritius and Tanzania), Asia (Hong Kong, Java, Nepal and Taiwan) and Europe (Italy). Deighton (1983) briefly described and discussed this species under the name *Cercoseptoria theae*, mainly based on Cavara’s original description, and reallocated it later to *Pseudocercospora* (Deighton 1987). Hsieh & Goh (1990: 327 and 328, Fig. 248) described and illustrated material of *P. theae* from Taiwan. *P. theae* is morphologically close to and confusable with *P. ocellata*. Several additional cercosporoid hyphomycetes are known as pathogens of *Camellia* spp. and have often been confused:

**Pseudocercospora camelliae** (Deighton) U. Braun (Braun 1993: 282)
≡ *Mycocentrospora camelliae* Deighton (Deighton 1983: 6).

On *Camellia* spp. (ornamental species), North America (USA, Georgia), New Zealand.

**Pseudocercospora camelliiicola** U. Braun & C.F. Hill (Braun & Hill 2002: 222)
≡ *Cercostigmina theae* (Deighton) U. Braun (Braun 1993: 108).

**Pseudocercospora javanica** Deighton (Deighton 1983: 5).

On *Camellia sinensis* var. *assamica* (J.W. Mast.) Kitam., Asia (Java).

**Pseudocercospora ocellata** (Deighton)

Deighton (Deighton 1987: 390)  
≡ *Cercoseptoria ocellata* Deighton (Deighton 1983: 2).  
≡ *Cercospora theae* Breda de Haan (Breda de Haan 1900: 11), non *Pseudocercospora theae* (Cavara) Deighton, 1987.

On *Camellia sinensis*, Africa (Nigeria and Tanzania), Asia (India, Indonesia, Japan and Sri Lanka) [Crous & Braun (2003)]. Recorded from India by Kamal (2010) and from Japan by Kobayashi (2007).

**Key to the species of Pseudocercospora on Camellia spp.**

1. Stromata lacking or small; mycelium internal and external; conidiophores in small fascicles, emerging through stromata, and solitary, arising from superficial hyphae; conidia acicular, 90–150 × 3–4 µm; on *Camellia* spp. in North America and New Zealand ............................................ *P. camelliae*

1* Stromata well-developed; superficial mycelium and solitary conidiophores lacking .. 2

2. Conidiomata sporodochial, composed of large immersed stromata, 50–120 µm diam., and numerous densely arranged conidiophores (stromata sometimes becoming spermatogonia or pycnidia with a single locule); conidiogenous cells unilocular, determinate or percurrently proliferating, annelations not very conspicuous; conidia ± acicular, 50–170(–190) × 2.5–4 µm; on *Camellia* spp. in Africa, Asia and New Zealand ................................. *P. camelliicola*

2* Stromata smaller, < 60 µm; conidiogenous cells with a single to several loci, proliferation sympodial; conidia somewhat shorter, about 30–125 µm ........................................ 3

3. Conidiophores relatively short, 5–40 × 1.5–4 µm, 0–2-septate, unbranched; conidia acicular to narrowly obclavate, base truncate to distinctly obconically truncate; on *Camellia* spp., Africa, Asia, Europe, South America ............................... *P. theae*

3* Conidiophores longer, 25–85 µm, up to 5-septate, occasionally branched; conidial base uniformly truncate or short obconically truncate ........................................... 4

4. Leaf spots small, up to about 2 mm diam., evident as typical eye-spots (at first black or purplish, later with pale center surrounded by a narrow swollen purplish brown margin); conidia acicular-filiform, base truncate, about 50–80 × 2–4 µm; on *Camellia sinensis*, Africa and Asia .................................................. *P. ocellata*

4* Leaf spots greyish brown, size and shape variable, up to 20 mm diam., margin indefinite; conidia narrowly obclavate-filiform, base short obconically truncate, longer, about 70–125 × 2.5–3 µm; on *Camellia sinensis*, only known from Java ............................................ *P. javanica*

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